DetectX®

CORTICOSTERONE Enzyme Immunoassay Kit

1 Plate Kit Catalog Number K014-H1
5 Plate Kit Catalog Number K014-H5

Species Independent

Sample Types Validated:

Serum, EDTA and Heparin Plasma, Urine, Dried Fecal Extracts, and Tissue Culture Media

Please read this insert completely prior to using the product. For research use only. Not for use in diagnostic procedures.

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# TABLE OF CONTENTS

- Background 3
- Assay Principle 4
- Related Products 4
- Supplied Components 5
- Storage Instructions 5
- Other Materials Required 6
- Precautions 6
- Sample Types 7
- Sample Preparation 7
- Reagent Preparation 8
- Assay Protocol 9
- Calculation of Results 10
- Typical Data 10-11
- Validation Data Sensitivity, Linearity, etc. 11-13
- Samples Values and Cross Reactivity 14
- Warranty & Contact Information 15
- Plate Layout Sheet 16
BACKGROUND

Corticosterone (C$_{21}$H$_{30}$O$_{4}$, Kendall’s Compound ‘B’) is a glucocorticoid secreted by the cortex of the adrenal gland. Corticosterone is produced in response to stimulation of the adrenal cortex by ACTH and is the precursor of aldosterone. Corticosterone is a major indicator of stress and is the major stress steroid produced in non-human mammals. Studies involving corticosterone and levels of stress include impairment of long term memory retrieval$^1$, chronic corticosterone elevation due to dietary restrictions$^2$ and in response to burn injuries$^3$. In addition to stress levels, corticosterone is believed to play a decisive role in sleep-wake patterns$^{4,5}$.

ASSAY PRINCIPLE

The DetectX® Corticosterone Immunoassay kit is designed to quantitatively measure Corticosterone present in serum, plasma, urine, extracted dried fecal samples, and tissue culture media samples. Please read the complete kit insert before performing this assay. This kit measures total corticosterone in serum and plasma and in extracted fecal samples.

A corticosterone stock solution is provided to generate a standard curve for the assay and all samples should be read off the standard curve. We provide protocols on page 8 to prepare assay standards from 5,000 to 78.125 pg/mL or from 10,000 to 78.125 pg/mL. Please choose the standard range that fits your sample concentrations most appropriately.

Standards or diluted samples are pipetted into a clear microtiter plate coated with an antibody to capture sheep antibodies. A corticosterone-peroxidase conjugate is added to the standards and samples in the wells. The binding reaction is initiated by the addition of a polyclonal antibody to corticosterone to each well. After an hour incubation the plate is washed and substrate is added. The substrate reacts with the bound corticosterone-peroxidase conjugate. After a short incubation, the reaction is stopped and the intensity of the generated color is detected in a microtiter plate reader capable of measuring 450nm wavelength. The concentration of the corticosterone in the sample is calculated, after making suitable correction for the dilution of the sample, using software available with most plate readers.

RELATED PRODUCTS

<table>
<thead>
<tr>
<th>Kits</th>
<th>Catalog No.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Urinary Creatinine Detection Kits</td>
<td>K002-H1/H5</td>
</tr>
<tr>
<td>Corticosterone Chemiluminescent Immunoassay Kits</td>
<td>K014-C1/C5</td>
</tr>
<tr>
<td>Cortisol Enzyme Immunoassay Kits (Strip Wells)</td>
<td>K003-H1/H5</td>
</tr>
<tr>
<td>Cortisol Enzyme Immunoassay Kits (Whole Plate)</td>
<td>K003-H1W/H5W</td>
</tr>
<tr>
<td>Cortisone Enzyme Immunoassay Kits</td>
<td>K017-H1/H5</td>
</tr>
<tr>
<td>Cortisone Chemiluminescent Immunoassay Kits</td>
<td>K017-C1/C5</td>
</tr>
</tbody>
</table>
SUPPLIED COMPONENTS

Coated Clear 96 Well Plates
A clear plastic microtiter plate(s) coated with donkey anti-sheep IgG.
Kit K014-H1 or -H5 1 or 5 Each Catalog Number X061-1EA

Corticosterone Standard
Corticosterone at 100,000 pg/mL in a special stabilizing solution.
Kit K014-H1 or -H5 125 µL or 625 µL Catalog Number C151-125UL or -625UL

DetectX® Corticosterone Antibody
A sheep polyclonal antibody specific for corticosterone.
Kit K014-H1 or -H5 3 mL or 13 mL Catalog Number C044-3ML or -13ML

DetectX® Corticosterone Conjugate
A corticosterone-peroxidase conjugate in a special stabilizing solution.
Kit K014-H1 or -H5 3 mL or 13 mL Catalog Number C045-3ML or -13ML

Assay Buffer Concentrate
A 5X concentrate that must be diluted with deionized or distilled water.
Kit K014-H1 or -H5 28 mL or 55 mL Catalog Number X065-28ML or -55ML

Dissociation Reagent
Kit K014-H1 or -H5 1 mL or 5 mL Catalog Number X058-1ML or -5ML
Dissociation Reagent is to be used only with Serum and Plasma samples.

Wash Buffer Concentrate
A 20X concentrate that should be diluted with deionized or distilled water.
Kit K014-H1 or -H5 30 mL or 125 mL Catalog Number X007-30ML or -125ML

TMB Substrate
Kit K014-H1 or -H5 11 mL or 55 mL Catalog Number X019-11ML or -55ML

Stop Solution
A 1M solution of hydrochloric acid. CAUSTIC.
Kit K014-H1 or -H5 5 mL or 25 mL Catalog Number X020-5ML or -25ML

Plate Sealer
Kit K014-H1 or -H5 1 or 5 Each Catalog Number X002-1EA

STORAGE INSTRUCTIONS
All components of this kit should be stored at 4°C until the expiration date of the kit.
OTHER MATERIALS REQUIRED

Distilled or deionized water.
Repeater pipet with disposable tips capable of dispensing 25 µL, 50 µL and 100 µL.
Colorimetric 96 well microplate reader capable of reading optical density at 450 nm.
Software for converting raw relative optical density readings from the plate reader and carrying out four parameter logistic curve (4PLC) fitting. Contact your plate reader manufacturer for details.

PRECAUTIONS

As with all such products, this kit should only be used by qualified personnel who have had laboratory safety instruction. The complete insert should be read and understood before attempting to use the product.

The antibody coated plate needs to be stored desiccated. The silica gel pack included in the foil ziploc bag will keep the plate dry. The silica gel pack will turn from blue to pink if the ziploc has not been closed properly.

This kit utilizes a peroxidase-based readout system. Buffers, including other manufacturers Wash Buffers, containing sodium azide will inhibit color production from the enzyme. Make sure all buffers used for samples are azide free. Ensure that any plate washing system is rinsed well with deionized water prior to using the supplied Wash Buffer as prepared on Page 8.

The Stop Solution is acid. The solution should not come in contact with skin or eyes. Take appropriate precautions when handling this reagent.
SAMPLE TYPES

This assay has been validated for serum, EDTA and heparin plasma, urine samples and for tissue culture samples. It has also been validated for dried fecal extract samples. Samples containing visible particulate should be centrifuged prior to using. Moderate to severely hemolyzed samples should not be used in this kit. Corticosterone can be assayed in other sample types by using one of the extraction protocols available on our website at: www.ArborAssays.com/resources/#protocols.

Corticosterone is identical across all species and we expect this kit may measure corticosterone from sources other than human. The end user should evaluate recoveries of corticosterone in other samples being tested.

SAMPLE PREPARATION

Serum and plasma samples need to be treated with the supplied Dissociation Reagent. Addition of this reagent will yield the total corticosterone concentration in serum or plasma. Dissociation Reagent is to be used only with Serum and Plasma samples.

Serum and Plasma Samples
Allow the Dissociation Reagent to warm completely to Room Temperature before use. We suggest pipetting 5 µL of Dissociation Reagent into 1 mL Eppendorf tubes. Add 5 µL of serum or plasma to the Dissociation Reagent in the tube, vortex gently and incubate at room temperature for 5 minutes or longer. Dilute with 490 µL of diluted Assay Buffer. This 1:100 dilution can be diluted further with diluted Assay Buffer. Final serum and plasma dilutions should be ≥ 1:100.

NOTE: Dissociation Reagent is to be used only with Serum and Plasma samples.

Urine Samples
Urine samples should be diluted ≥ 1:20 with the diluted Assay Buffer prior running in the assay. Please see our Urinary Creatinine Detection kits, K002-H1 and K002-H5, for assays to measure urine creatinine which can be used to allow normalization of corticosterone in a random urine specimen.

Dried Fecal Samples:
We have a detailed Extraction Protocol available on our website at: www.ArborAssays.com/resources/#protocols. The ethanol concentration in the final diluted Assay Buffer dilution added to the well should be <5%.

Tissue Culture Media
For measuring corticosterone in tissue culture media (TCM), samples should be read off a standard curve generated in TCM. Samples may need to be diluted further in TCM. We have validated the assay using RPMI-1640.

Use all Samples within 2 hours of preparation, or stored at ≤ -20°C until assaying.
REAGENT PREPARATION

Allow the kit reagents to come to room temperature for 30 minutes. Ensure that all samples have reached room temperature and have been diluted as appropriate prior to running them in the kit.

Assay Buffer
Dilute Assay Buffer Concentrate 1:5 by adding one part of the concentrate to four parts of deionized water. Once diluted this is stable for 3 months at 4°C.

Wash Buffer
Dilute Wash Buffer Concentrate 1:20 by adding one part of the concentrate to nineteen parts of deionized water. Once diluted this is stable for 3 months at room temperature.

Standard Preparation
Label test tubes as #1 through #8. Pipet 450 µL of Assay Buffer into tube #1 and 250 µL into tubes #2 to #8. The corticosterone stock solution contains an organic solvent. Prerinse the pipet tip several times to ensure accurate delivery. Carefully add 50 µL of the corticosterone stock solution to tube #1 and vortex completely. Take 250 µL of the corticosterone solution in tube #1 and add it to tube #2 and vortex completely. Repeat the serial dilutions for tubes #3 through #8. The concentration of corticosterone in tubes 1 through 8 will be 10,000, 5,000, 2,500, 1,250, 625, 312.5, 156.25, and 78.125 pg/mL.

Use all Standards within 2 hour of preparation.

<table>
<thead>
<tr>
<th>Std 1</th>
<th>Std 2</th>
<th>Std 3</th>
<th>Std 4</th>
<th>Std 5</th>
<th>Std 6</th>
<th>Std 7</th>
<th>Std 8</th>
</tr>
</thead>
<tbody>
<tr>
<td>Assay Buffer (µL)</td>
<td>450</td>
<td>250</td>
<td>250</td>
<td>250</td>
<td>250</td>
<td>250</td>
<td>250</td>
</tr>
<tr>
<td>Addition Stock</td>
<td>Std 1</td>
<td>Std 2</td>
<td>Std 3</td>
<td>Std 4</td>
<td>Std 5</td>
<td>Std 6</td>
<td>Std 7</td>
</tr>
<tr>
<td>Vol of Addition (µL)</td>
<td>50</td>
<td>250</td>
<td>250</td>
<td>250</td>
<td>250</td>
<td>250</td>
<td>250</td>
</tr>
<tr>
<td>Final Conc (pg/mL)</td>
<td>10,000</td>
<td>5,000</td>
<td>2,500</td>
<td>1,250</td>
<td>625</td>
<td>312.5</td>
<td>156.25</td>
</tr>
</tbody>
</table>
ASSAY PROTOCOL

We recommend that all standards and samples be run in duplicate to allow the end user to accurately determine corticosterone concentrations.

1. Use the plate layout sheet on the back page to aid in proper sample and standard identification. Determine the number of wells to be used and return unused wells to the foil pouch with desiccant. Seal the ziploc plate bag and store at 4°C.

2. Pipet 50 μL of samples or standards into wells in the plate.

3. Pipet 75 μL of Assay Buffer into the non-specific binding (NSB) wells.

4. Pipet 50 μL of Assay Buffer into the maximum binding (B0 or Zero standard) wells.

5. Add 25 μL of the DetectX® Corticosterone Conjugate to each well using a repeater pipet.

6. Add 25 μL of the DetectX® Corticosterone Antibody to each well, except the NSB wells, using a repeater pipet.

7. Gently tap the sides of the plate to ensure adequate mixing of the reagents. Cover the plate with the plate sealer and shake at room temperature for 1 hour. If the plate is not shaken signals bound will be approximately 45% lower.

8. Aspirate the plate and wash each well 4 times with 300 μL wash buffer. Tap the plate dry on clean absorbent towels.

9. Add 100 μL of the TMB Substrate to each well, using a repeater pipet.

10. Incubate the plate at room temperature for 30 minutes without shaking.

11. Add 50 μL of the Stop Solution to each well, using a repeater or a multichannel pipet.

12. Read the optical density generated from each well in a plate reader capable of reading at 450 nm.

13. Use the plate reader’s built-in 4PLC software capabilities to calculate corticosterone concentration for each sample.

NOTE: If you are using only part of a strip well plate, at the end of the assay throw away the used wells and retain the plate frame for use with the remaining unused wells.
CALCULATION OF RESULTS

Average the duplicate OD readings for each standard and sample. Create a standard curve by reducing the data using the 4PLC fitting routine on the plate reader, after subtracting the mean OD's for the NSB. The sample concentrations obtained, calculated from the %B/B0 curve, should be multiplied by the dilution factor to obtain neat sample values.

Or use the online tool from MyAssays to calculate the data: www.myassays.com/arbor-assays-corticosterone-enzyme-immunoassay-kit.assay

<table>
<thead>
<tr>
<th>Sample</th>
<th>Mean OD</th>
<th>Net OD</th>
<th>% B/B0</th>
<th>Corticosterone Conc. (pg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NSB</td>
<td>0.067</td>
<td>0</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Standard 1</td>
<td>0.242</td>
<td>0.175</td>
<td>11.9</td>
<td>10,000</td>
</tr>
<tr>
<td>Standard 2</td>
<td>0.330</td>
<td>0.263</td>
<td>17.9</td>
<td>5,000</td>
</tr>
<tr>
<td>Standard 3</td>
<td>0.478</td>
<td>0.411</td>
<td>28.0</td>
<td>2,500</td>
</tr>
<tr>
<td>Standard 4</td>
<td>0.656</td>
<td>0.589</td>
<td>40.1</td>
<td>1,250</td>
</tr>
<tr>
<td>Standard 5</td>
<td>0.876</td>
<td>0.809</td>
<td>55.1</td>
<td>625</td>
</tr>
<tr>
<td>Standard 6</td>
<td>1.070</td>
<td>1.003</td>
<td>68.3</td>
<td>312.5</td>
</tr>
<tr>
<td>Standard 7</td>
<td>1.269</td>
<td>1.202</td>
<td>81.8</td>
<td>156.25</td>
</tr>
<tr>
<td>Standard 8</td>
<td>1.400</td>
<td>1.333</td>
<td>90.7</td>
<td>78.125</td>
</tr>
<tr>
<td>B0</td>
<td>1.536</td>
<td>1.469</td>
<td>100.0</td>
<td>0</td>
</tr>
<tr>
<td>Sample 1</td>
<td>0.442</td>
<td>0.375</td>
<td>25.5</td>
<td>2,894</td>
</tr>
<tr>
<td>Sample 2</td>
<td>1.109</td>
<td>1.042</td>
<td>70.9</td>
<td>283.2</td>
</tr>
</tbody>
</table>

Always run your own standard curve for calculation of results. Do not use this data.

Conversion Factor: 100 pg/mL of corticosterone is equivalent to 288.6 pM.
Typical Normal Range Standard Curves

Always run your own standard curves for calculation of results. Do not use this data.

VALIDATION DATA

Sensitivity and Limit of Detection
Sensitivity was calculated by comparing the OD’s for nineteen wells run for each of the B0 and standard #7. The detection limit was determined at two (2) standard deviations from the B0 along the standard curve. Sensitivity was determined as 18.6 pg/mL.

The Limit of Detection for the assay was determined in a similar manner by comparing the OD’s for twenty runs for each of the zero standard and a low concentration human sample. Limit of Detection was determined as 16.9 pg/mL.
Linearity
Linearity was determined by taking two serum samples treated with Dissociation Reagent and diluted 1:50 with Assay Buffer, one with a low diluted corticosterone level of 104.6 pg/mL and one with a higher diluted level of 2,890.5 pg/mL, and mixing them in the ratios given below. The measured concentrations were compared to the expected values based on the ratios used.

<table>
<thead>
<tr>
<th>Low Serum</th>
<th>High Serum</th>
<th>Expected Conc. (pg/mL)</th>
<th>Observed Conc. (pg/mL)</th>
<th>% Recovery</th>
</tr>
</thead>
<tbody>
<tr>
<td>80%</td>
<td>20%</td>
<td>661.8</td>
<td>654.0</td>
<td>98.8</td>
</tr>
<tr>
<td>60%</td>
<td>40%</td>
<td>1,219.0</td>
<td>1,232.3</td>
<td>101.1</td>
</tr>
<tr>
<td>40%</td>
<td>60%</td>
<td>1,776.1</td>
<td>1,763.9</td>
<td>99.3</td>
</tr>
<tr>
<td>20%</td>
<td>80%</td>
<td>2,333.3</td>
<td>2,249.5</td>
<td>96.4</td>
</tr>
</tbody>
</table>

Mean Recovery 98.9%

Linearity

\[
y = 0.9545x + 45.521
\]

\[R^2 = 0.9985\]
**Intra Assay Precision**
Four human samples were diluted with Assay Buffer and run in replicates of 20 in an assay. The mean and precision of the calculated Corticosterone concentrations were:

<table>
<thead>
<tr>
<th>Sample</th>
<th>Corticosterone Conc. (pg/mL)</th>
<th>%Cv</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>2,460.6</td>
<td>6.3</td>
</tr>
<tr>
<td>2</td>
<td>601.5</td>
<td>6.5</td>
</tr>
<tr>
<td>3</td>
<td>371.6</td>
<td>3.1</td>
</tr>
<tr>
<td>4</td>
<td>259.0</td>
<td>4.8</td>
</tr>
</tbody>
</table>

**Inter Assay Precision**
Three human samples were diluted with Assay Buffer and run in duplicates in fourteen assays run over multiple days by four operators. The mean and precision of the calculated Corticosterone concentrations were:

<table>
<thead>
<tr>
<th>Sample</th>
<th>Corticosterone Conc. (pg/mL)</th>
<th>%CV</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>2,618.3</td>
<td>7.5</td>
</tr>
<tr>
<td>2</td>
<td>630.1</td>
<td>6.4</td>
</tr>
<tr>
<td>3</td>
<td>267.9</td>
<td>9.9</td>
</tr>
</tbody>
</table>
SAMPLE VALUES

Six random mammalian serum and plasma samples were tested in the assay. Neat sample values ranged from 0.87 to 38.5 µg/dL with an average for the human samples of 1.56 µg/dL. The normal reference range for serum corticosterone is 0.13-2.3 µg/dL6.

Dried fecal samples were processed as described on page 7 and run in the assay. Samples kindly donated by Dr. J. Williams at the Indianapolis Zoo, which included Amur Tiger, Giraffe, Kudu, Lion, Reeves Muntjac, White Handed Gibbon, White Rhino, and Zebra, were tested and corticosterone values obtained ranged from 7.85 to 81.6 pg/mg dried fecal material.

Palme and Möestl and colleagues have shown that radiolabeled administered glucocorticoids are excreted in differing amounts in urine and feces7 across species, with fecal excretion ranging from 7% of administered cortisol in the pig to 82% in the cat8-10. Palme has also shown that the peak of fecal glucocorticoid concentrations occur at 12 hours for sheep, but takes 48 hours to peak in pigs. It is therefore necessary to evaluate the timing and relative fecal or urine excretion of glucocorticoids for each species.


CROSS REACTIVITY

The following cross reactants were tested in the assay and calculated at the 50% binding point.

<table>
<thead>
<tr>
<th>Steroid</th>
<th>Cross Reactivity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Corticosterone</td>
<td>100%</td>
</tr>
<tr>
<td>Desoxycorticosterone</td>
<td>12.30%</td>
</tr>
<tr>
<td>Tetrahydrocorticosterone</td>
<td>0.76%</td>
</tr>
<tr>
<td>Aldosterone</td>
<td>0.62%</td>
</tr>
<tr>
<td>Cortisol</td>
<td>0.38%</td>
</tr>
<tr>
<td>Progesterone</td>
<td>0.24%</td>
</tr>
<tr>
<td>Dexamethasone</td>
<td>0.12%</td>
</tr>
<tr>
<td>Corticosterone-21-Hemisuccinate</td>
<td>&lt; 0.1%</td>
</tr>
<tr>
<td>Cortisone</td>
<td>&lt; 0.08%</td>
</tr>
<tr>
<td>Estradiol</td>
<td>&lt; 0.08%</td>
</tr>
</tbody>
</table>
LIMITED WARRANTY

Arbor Assays warrants that at the time of shipment this product is free from defects in materials and workmanship. This warranty is in lieu of any other warranty expressed or implied, including but not limited to, any implied warranty of merchantability or fitness for a particular purpose.

We must be notified of any breach of this warranty within 48 hours of receipt of the product. No claim shall be honored if we are not notified within this time period, or if the product has been stored in any way other than outlined in this publication. The sole and exclusive remedy of the customer for any liability based upon this warranty is limited to the replacement of the product, or refund of the invoice price of the goods.

CONTACT INFORMATION

For details concerning this kit or to order any of our products please contact us:

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- Contracts@ArborAssays.com

OFFICIAL SUPPLIER TO ISWE

Arbor Assays and the International Society of Wildlife Endocrinology (ISWE) signed an exclusive agreement for Arbor Assays to supply ISWE members with EIA kits for wildlife conservation research.